

REMARKS

The substitute specification has been submitted. Claims 5, 13, 14, 17-26 and 28 have been cancelled. Claims 1-4, 6-12, 15, 16 and 27 remain in the application. Each claim has been amended. Amendments to Figure 1 are submitted for the Examiner's approval. Corrected Figures 2, 3 and 4 are submitted. Reexamination and reconsideration of the application are requested.

Discussion of Amendments

Substitute Specification

The original specification contained numerous typographical errors. In particular, kringle 1-3 region was erroneously referred to as kringle 1-5 region throughout the application, including the title and the abstract, as a result of an unintentional clerical error. This has been amended throughout the substitute specification. Support for these amendments is found in the Examples and Figures 2-4, which illustrate the purification and antiangiogenic properties of kringle 1-3 region fragment of plasminogen. Additionally, Figure 1 and the Sequence Listing (SEQ ID NO:1 and SEQ ID NO:2) both disclose the amino acid and nucleotide sequence of kringle 1-3 region of plasminogen. Accordingly, the specification has been substituted to correctly identify the deglycosylated kringle region fragment of plasminogen and its antiangiogenic properties as "kringle 1-3" region fragment, instead of incorrectly referring to it as "kringle 1-5" region fragment. In view of these amendments, on page 13, lines 17-19 were amended to recite "It is to be understood that "kringle 1-3 region" is defined herein as a region that corresponds to approximately amino acid position 1 through approximately amino acid position 333 of human plasminogen.", instead of the previously identified borders of the kringle 1-5 region fragment. Both a marked-up copy of the specification to show the changes made, and a clean copy of the amended specification are included with this response. In the marked-up copy, inserted text is marked by underline (inserted text), and deleted text is marked by strikethrough (~~deleted text~~). Changed lines are indicated by a vertical line at the right border of the text.

Additional corrections to the application are discussed in more detail below. These include the removal of hyperlinks and corrections to Figures 1-4.

Claims 1 and 27 have been amended to recite "kringle 1-3 region fragment of a plasminogen protein" in order to more distinctly claim the invention. Claim 1 was also amended to clarify that it refers to the composition comprising deglycosylated form of kringle 1-3 region fragment of plasminogen and, optionally, a glycosylated form of kringle 1-3 region fragment of plasminogen. Claims 4 and 9 has been amended to recite "amino acid position corresponding to the N-glycosylation site of human plasminogen of human plasminogen" in order to more distinctly claim the invention. Claim 5 was canceled as redundant. Dependent Claims 2-4, 6-12, and 15-16 have been amended to recite kringle 1-3 region fragment.

1. Election/Restriction

2. Claims 13, 14, 17-26 and 28 are drawn to non-elected claims. Applicants have cancelled Claims 13, 14, 17-26 and 28.

3. Priority

The Office Action has rejected the claim for domestic priority to U.S. Provisional Patent Application Serial No. 60/119,562, filed February 10, 1999 and U.S. Provisional Patent Application Serial No. 60/128,062, filed April 7, 1999 because the Provisional Patent Applications do not disclose the use of Kringle 1-5.

This rejection is respectfully traversed. The Specification has been amended to replace K1-5 with K1-3 throughout the document. Support for these amendments is discussed earlier.

Additionally, U.S. Provisional Patent Application Serial No. 60/119,562 discloses the purification and use of same kringle region fragment of human plasminogen as the present application. In particular, Example 1 in the section "Detailed Description" of the present application corresponds to the sections "Proteins", "Generation of angiostatin", "Bovine Endothelial Cell Proliferation assay" and "Statistical Analysis" on pages 5-6 of the Provisional Patent Application Serial No. 60/119,562. Example 2 in the section "Detailed Description" of the present application corresponds to the section "Generation of angiostatin from plasminogen 1 and plasminogen 2 by porcine pancreatic elastase" on page 7 of the Provisional Patent Application. Example 3 of the present application corresponds to the section "Inhibition of bovine endothelial cell proliferation by angiostatin 1 and 2" on pages 7 and 8 of the Provisional Patent Application. Figures 2, 3 and 4 of the present application are substantially identical to Figures 1a, 1b and 3 of the Provisional Patent Application, and illustrate the same experimental

data regarding the isolation and antiangiogenic properties of the glycosylated and deglycosylated forms of the same kringle region fragment of human plasminogen. The two glycoforms of this fragment are characterized by their respective molecular mass of 40 kDa and 38kDa determined by SDS-PAGE, as described on page 7 of the Provisional Patent Application and page 37, lines 10-11 of the present application. Antiangiogenic properties of this deglycosylated kringle region fragment of human plasminogen and the idea that deglycosylation modifies the such properties are further described on pages 9 and 10 of the Provisional Patent Application.

Based on its molecular weight of 38-40 kDa, the disclosed kringle region fragment, should be correctly identified as kringle 1-3 region fragment. However, due to a clerical error it was incorrectly referred to in the present application as kringle 1-5 region fragment, which would have an estimated molecular weight of at least 50kDa. The amendment of the specification to recite kringle 1-3 region fragment is believed to correct this discrepancy.

In view of the amendments, it should be clear that U.S. Provisional Patent Application Serial No. 60/119,562 discloses the same subject matter as the present application and provides support for all of the claims of the application. Therefore, the Applicants respectfully request the withdrawal of the objection to the claim of priority to U.S. Provisional Patent Application Serial No. 60/119,562.

The rejection of the claim for domestic priority to U.S. Provisional Patent Application Serial No. 60/128,062, filed April 7, 1999 is respectfully traversed. U.S. Provisional Patent Application Serial No. 60/128,062 discloses that the tumor suppressing activity of angiostatin protein resides with kringles 1 to 3. Applicants consider this a subject matter of their invention as disclosed in the amended specification and claimed in Claims 1-4, Claims 6-12, Claims 15-16, and Claim 27 of the present application. The Applicants respectfully request the withdrawal of the objection to the claim of priority to U.S. Provisional Patent Application Serial No. 60/128,06.

4. Drawings

The Office Action has objected to the drawings. Figures 2, 3, and 4 are rejected by the Draftsperson under 37 CFR 1.84. The Applicants has corrected the drawings as required by the Office Action.

Figures 2 and 3 were corrected to contain acceptable right and left margins pursuant to 37 CFR 1.84(g). Figures 2-4 were corrected to show uniformly thick and well-defined, clean, durable and black lines, and plain and legible numbers and reference characters pursuant to 37 CFR 1.84(i and p).

Specification

5. The Office Action has objected to the disclosure because it contains an embedded hyperlink and/or other form of browser-executable code. The Applicants have deleted the embedded hyperlinks in the substitute specification. The specification is now believed to comply with MPEP §608.01.

6. The Office Action has objected to the disclosure of the invention because of the discrepancies between Figure 1 and its brief description. In particular, the sequence identification numbers (ID) SEQ ID NO:1 and SEQ ID NO:2 for nucleotide and amino acid sequences, respectively, of kringle 1-3 region fragments of human plasminogen do not agree.

Throughout the substitute specification, the Applicants refer to nucleotide sequence as SEQ ID NO:1 and amino acid sequence as SEQ ID NO:2 to correspond to the sequence identification numbers assigned by the "Patentin" software, ver. 3.0 used in the preparation of the formal Sequence Listing. In the initially submitted application, amino acid sequence was referred to as SEQ ID NO:1, and nucleotide sequence was referred to as SEQ ID NO:2 both in the specification and in Figure 1. A Preliminary Amendment submitted by the Applicant on February 16, 2001, states that in preparing the Sequence Listing, Applicants used "Patentin" Software, ver. 3.0, which required the nucleotide sequence to be SEQ ID NO:1, and the amino acid sequence to be SEQ ID NO:2. Therefore, Applicants requested a global change in the sequence identification numbers so that SEQ ID NO:1 became SEQ ID NO:2, and that SEQ ID NO:2 became SEQ ID NO:1 throughout the Specification, including the Claims. Figure 1, however, was not corrected and continued to show amino acid sequence as SEQ ID NO:1, and

nucleotide sequence as SEQ ID NO:2, which created the discrepancy corrected by the present amendment to Figure 1.

Applicants therefore submit amendments to Figure 1 (in red) for the Examiner's review. The amendments correct sequence identification number SEQ ID NO:1 for the amino acid sequence to SEQ ID NO:2, and SEQ ID NO:2 for the nucleotide sequence to SEQ ID NO:1, to make the sequence ID numbers consistent with the specification and the Sequence Listing. Applicants have reviewed the entire specification for similar errors. Applicants submit that the amendment corrects the informality.

7. Claim Rejections under 35 USC §112

8. The Office Action has rejected Claims 1-12, 15, 16, and 27 under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

a. The Office Action has rejected Claims 1 and 27 as vague and indefinite in the recitation of the "fragment of a kringle 1-5 region of plasminogen". The Office Action contends that it is not clear as to what kringles should comprise the fragment: all of the kringles, 1-5, or 1-3, and, therefore, the metes and bounds of the claims cannot be determined.

Claims 1 and 27 have been amended to recite "kringle 1-3 region fragment of a plasminogen protein". Support for this amendment is found throughout the amended specification. The rejection to Claim 1 is now believed to be overcome.

Claim 5 was cancelled as redundant upon amendment of Claim 1. Dependent Claims 2-4, 6-12, and 15-16 have been amended to recite kringle 1-3 region fragment in agreement with Claim 1.

Claim 27, in its amended form it recites: "A deglycosylated kringle 1-3 region fragment of a plasminogen protein, wherein the deglycosylated fragment amino acid sequence is shown in SEQ ID NO:2." SEQ ID NO:2 is unambiguously identified throughout the amended application as that of kringle 1-3 region fragment of plasminogen. Support for this amendment is found throughout the amended specification. The rejection to Claim 27 is now believed to be overcome.

b. The Office Action has rejected Claim 1 as vague and indefinite because it was not clear whether or not the composition should comprise just the deglycosylated fragment or both the deglycosylated fragment and the naturally glycosylated form. Claim 1 was amended to clarify that it refers to the composition comprising deglycosylated form of kringle 1-3 region fragment of plasminogen and, optionally, a glycosylated form of kringle 1-3 region fragment of plasminogen. Support for these amendment can be found in on page 9, lines 20-24, and Example 3 of the written description of the invention. Therefore, the rejection is believed to be overcome.

c. The Office Action has rejected Claims 4 and 9 as vague and indefinite because these claims recite "amino acid position of Asn-289", whereas the amino acid sequence of SEQ ID NO:2 consist of 260 amino acid residues. Claims 4 and 9 has been amended to recite "amino acid position corresponding to the N-glycosylation site of human plasminogen" in order to more distinctly claim the invention. Support for these amendments can be found in on page 9, last paragraph, and page 10, first paragraph of the written description of the invention. Claims 4 and 9 in their amended form refer to the sequence of human plasminogen, which is readily available from public protein sequence databases, such as SWISS-PROT (<http://www.expasy.ch/sprot/sprot-top.html>, protein sequence Accession Number P00747). Human plasminogen is described in the written description of the invention at page 4, lines 17-19, page 11, lines 29-30, and page 37, lines 16-18. The description of the amino acid position described as "corresponding to the N-glycosylation site of human plasminogen" will be readily understood by those skilled in the art. It has long been known in the art that N-glycosylation occurs at a specific consensus sequence Asn-Xaa-Ser/Thr. There is only one such position in human plasminogen molecule, which occurs in the sequence Asn-Arg-Thr. Before the complete sequence of human plasminogen was known, the asparagine in this sequence was referred to as "Asn-289" (see, for example, Schaller, et al., Fibrinolysis, 1987, v. 1, pp. 91-102). Upon cloning of the complete plasminogen sequence, the sequence numbering was corrected and the glycosylation site is now referred to as "Asn-308" (see protein sequence Accession Number P00747 in the Swiss Prot database). The definition of "corresponding to " when referring to amino acids is provided on page 11, lines 16-20 of the written description of the invention. The objection is believed to be overcome.

9. Claim Rejections under 35 USC §102

10. The Office Action has rejected Claims 1-6, 8, 9 12, 15 and 16 under 35 USC 102(b) as being anticipated by Accession Number P00747 and Sim, *et al.* (Cancer Research

57:1329-1334, April 1, 1997). The rejection is respectfully traversed. Protein sequence Accession Number P00747 teaches amino acid sequence of full length human plasminogen along with its N-glycosylation site at Asn-308. Claim 1, on the other hand, is directed to a composition comprising a pharmaceutically acceptable carrier, deglycosylated kringle 1-3 region fragment, and, optionally, a glycosylated kringle 1-3 region fragment, not full length human plasminogen. Kringle region fragments, by definition, do not include the full length human plasminogen molecule. See, for example, page 3, lines 22-26 of the written description of the invention.

The Office Action contends that Sim, *et al.* teaches recombinant angiostatin protein that is approximately kringles 1-3 and the use of its deglycosylated form for proliferation of bovine capillary endothelial cells in vitro and suppression of the growth of Lewis lung carcinoma low metastatic (LLC-LM) phenotype metastases in C57BL/6 mice *in vivo*. Applicants respectfully bring to the Examiner's attention that on page 1331, column, Sim, *et al.* describes angiostatin protein that appeared on SDS-PAGE gel as a double band with an apparent molecular mass of 49 kDa and 51 kDa, which corresponds to kringle 1-4 region fragment of the plasminogen molecule. In the abstract of Sim, *et al.* the fragment is clearly identified as "a recombinant protein comprising kringles 1-4 of human plasminogen (amino acids 93-470)".

Moreover, Sim, *et al.* does not teach or suggest the use of **deglycosylated** or **unglycosylated** form of angiostatin protein for suppression of angiogenesis *in vitro* or *in vivo*. As noted by the Office Action, on column 2 on page 1331, Sim, *et al.* describes endoglycosidase H treatment of this angiostatin protein, which resulted in the disappearance of the double band and caused recombinant angiostatin to migrate at 51 kDa. This experimental data led to the conclusion that recombinant angiostatin used in the study exists in both N-glycosylated and unglycosylated form. However, the sentence prior to the description of these experiments (last sentence of column 1 on page 1331) notes that it was the "preparation of recombinant Angiostatin protein **consisting of the doublet** was used in **all *in vitro* and *in vivo* assays** reported in this paper". Thus, the use of **deglycosylated** or **unglycosylated** plasminogen fragments for inhibition of angiogenesis is not taught or suggested by Sim, *et al.*, and the objection should be withdrawn.

Version With Markings To Show Changes Made

Amendments In the Specification:

In accordance with 37 CFR 1.121(b), the following replacement paragraphs show all the changes made by the foregoing amendment relative to the previous version of the paragraphs.

The Specification:

A marked-up version of the amended Specification is attached. Inserted text is marked by underline (inserted text), and deleted text is marked by strikethrough (~~deleted text~~). Changed lines are indicated by a vertical line at the right border of the text.

In the Drawings:

Submitted for the Examiner's approval, the amendments to the to the Figure 1 are shown in red in compliance with 37 CFR 1.121(d).

In compliance with 37 CFR 1.85 submitted are corrected Figures 2, 3 and 4.

In the Claims:

In accordance with 37 CFR 1.121(c), the following versions of the claims as rewritten by the foregoing amendment show all the changes made relative to the previous versions of the claims.

Claims 13, 14, 17-26 and 28 have been cancelled.

Claims 1-4, 6-12, 15, 16, and 27 have been amended as follows (inserted text is marked by underline (inserted text), and deleted text is marked by strikethrough (~~deleted text~~)):

1. (Amended) A composition comprising a pharmaceutically acceptable carrier, and a deglycosylated kringle 1-3 region ~~fragment of a kringle 1-5 region~~ of a plasminogen protein,

and, optionally, a naturally glycosylated kringle 1-3 region fragment of a plasminogen protein, in a greater amount than a naturally glycosylated form of the deglycosylated fragment, wherein the deglycosylated kringle 1-3 region fragment lacks one or more carbohydrate moieties linked to the naturally glycosylated form of the fragment, and wherein the deglycosylated fragment has antiangiogenic activity, and wherein a naturally glycosylated kringle 1-3 region fragment, when present, is in a smaller amount than deglycosylated kringle 1-3 region fragment.

2. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment lacks a bisialylated-biantennary glycan.

3. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment lacks an N-linked carbohydrate moiety.

4. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment lacks a carbohydrate chain at amino acid position Asn-289 corresponding to the N-glycosylation site of human plasminogen.

6. (Amended) The composition of claim 5, wherein the deglycosylated kringle 1-3 region fragment begins at approximately amino acid 87 of human plasminogen.

7. (Amended) The composition of claim 5, wherein the deglycosylated kringle 1-3 region fragment amino acid sequence is shown in SEQ ID NO:2.

8. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment is produced recombinantly.

9. (Amended) The composition of ~~claim~~ claim 1, wherein the deglycosylated kringle 1-3 region fragment has an amino acid substitution at amino acid position ~~289~~ corresponding to the N-glycosylation site of human plasminogen.

10. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment and the glycosylated form of the fragment are at a ratio of at least 60:40.

11. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment and the glycosylated form of the fragment are at a ratio of at least 80:20.

12. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment and the glycosylated form of the fragment are at a ratio of 100:0.

15. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment has antiangiogenic activity in vitro.

16. (Amended) The composition of claim 1, wherein the deglycosylated kringle 1-3 region fragment has antiangiogenic activity in vivo.

27. (Amended) A deglycosylated kringle 1-3 region fragment of a ~~kringle 1-5 region~~ of a plasminogen protein, wherein the deglycosylated kringle 1-3 region fragment amino acid sequence is shown in SEQ ID NO:2.

11. Conclusion

For the reasons set forth above, the present application is believed to be in condition for allowance. Reconsideration of the rejections is requested, and allowance of the claims at an early date is courteously solicited.

If the Examiner believes any informalities remain in the application that may be corrected by Examiner's amendment, or there are any other issues that can be resolved by telephone interview, a telephone call to the undersigned attorney at (404) 745-2413.

No additional fees are believed due, however, the Commissioner is hereby authorized to charge any deficiencies which may be required or credit any overpayment to Deposit Account Number 11-0855.

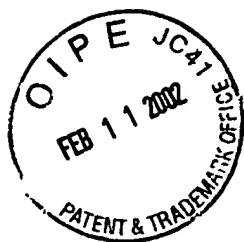
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**Version With Markings To Show Changes Made to
Substitute Specification**



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**DEGLYCOSYLATED KRINGLE 4-51-3 REGION
FRAGMENTS OF PLASMINOGEN
AND METHODS OF USE**

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This invention was made with government support under Grant No.(s) CA 45548 by the NIH. The government may have certain rights in the invention.

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Cross Reference to Related Applications

This application claims priority to U.S. Provisional Patent Application Serial No. 60/119,562, filed February 10, 1999 and U.S. Provisional Patent Application Serial No. 60/128,062, filed April 7, 1999.

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Field of the Invention

The present invention relates to compositions and methods for the modulation of angiogenesis. More particularly, the present invention relates to deglycosylated kringle 4-51-3 region proteins that are useful for the treatment of angiogenesis-associated diseases such as cancer.

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Background of the Invention

As used herein, the term "angiogenesis" means the generation of new blood vessels into a tissue or organ. Under normal physiological conditions, humans or animals undergo angiogenesis only in very specific restricted situations. For example, angiogenesis is normally observed in wound healing,

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fetal and embryonal development and formation of the corpus luteum, endometrium and placenta. However, angiogenesis also occurs under abnormal or undesired conditions such as during tumor development, growth and metastasis. This type of angiogenesis may also be referred to as uncontrolled angiogenesis.

Both controlled and uncontrolled angiogenesis are thought to proceed in a similar manner. Endothelial cells and pericytes surrounded by a basement membrane form capillary blood vessels. Angiogenesis begins with the erosion of the basement membrane by enzymes released by endothelial cells and leukocytes. The endothelial cells, which line the lumen of blood vessels, then protrude through the basement membrane. Angiogenic stimulants induce the endothelial cells to migrate through the eroded basement membrane. The migrating cells form a "sprout" off the parent blood vessel, where the endothelial cells undergo mitosis and proliferate. The endothelial sprouts merge with each other to form capillary loops, creating the new blood vessel.

Persistent, unregulated angiogenesis occurs in a multiplicity of disease states, tumor metastasis and abnormal growth by endothelial cells. The diverse pathological disease states in which unregulated angiogenesis is present have been grouped together as angiogenic dependent or angiogenic associated diseases. The hypothesis that tumor growth is angiogenesis-dependent was first proposed in 1971 by M. Judah Folkman. (Folkman J., "Tumor angiogenesis: Therapeutic implications" *N. Engl. Jour. Med.* 285:1182-1186 (1971)). In its simplest terms the hypothesis states: "Once tumor 'take' has occurred, every increase in tumor cell population must be preceded by an increase in new capillaries converging on the tumor." Tumor 'take' is currently understood to indicate a pre-vascular phase of tumor growth in which a population of tumor cells occupying a few cubic millimeters volume and not exceeding a few million cells, can survive on existing host

microvessels. Expansion of tumor volume beyond this phase requires the induction of new capillary blood vessels.

Several molecules have been discovered that inhibit angiogenesis, inhibit tumor growth, cause regression of primary tumors, and/or inhibit metastasis of primary tumors. These molecules are called antiangiogenic agents. One of these antiangiogenic agents has been termed angiostatin, which is a fragment of a plasminogen protein. Angiostatin was first described in U.S. Patent No. 5,639,725.

Plasminogen protein comprises five kringle region domains and a serine protease domain located at the carboxy-terminal region. The DNA sequence of human plasminogen has been published. (Browne, M.J., *et al.*, "Expression of recombinant human plasminogen and aglycoplasminogen in HeLa cells" *Fibrinolysis* 5(4):257-260 (1991)). Each kringle region of the plasminogen molecule contains approximately 80 amino acids and contains 3 disulfide bonds. (Robbins, K.C., "The plasminogen-plasmin enzyme system" *Hemostasis and Thrombosis, Basic Principles and Practice*, 2nd Edition, ed. by Colman, R.W. *et al.* J.B. Lippincott Company, pp. 340-357, 1987). The approximate amino acid spans of each kringle domain of human plasminogen are as follows: kringle 1 typically encompasses Cys84-Cys162, kringle 2 typically encompasses Cys166-Cys243, kringle 3 typically encompasses Cys256-Cys333, kringle 4 typically encompasses Cys358-Cys435, and kringle 5 typically encompasses Cys462-Cys541. (Castellino, F.J. and McCance, S.G., "The kringle domains of human plasminogen" *Ciba Found. Symp.*, 212:46-65 (1997)). Because each kringle domain is separated by an approximately 20 amino acid inter-kringle domain, a kringle region as herein defined can include any portion of these adjacent inter-kringle domains.

Angiostatin protein comprises one or more of these five kringle regions of plasminogen. Among the proteases suggested to be responsible for angiostatin generation are

macrophage metalloelastase, metalloproteinases (mmp's) -3, - 7, and -9 and plasmin itself in the presence of a free sulphhydryl donor such as cysteine. (Dong, Z. *et al.* "Macrophage-derived metalloelastase is responsible for the generation of angiostatin in Lewis lung carcinoma" *Cell*, 88:801-810 (1997); Lijnen, H.R. *et al.*, "Generation of an angiostatin-like fragment from plasminogen by stromelysin-1 (MMP-3)" *Biochemistry*, 37:4699-4702 (1998); Patterson, B.C. and Sang Q.A., "Angiostatin-converting enzyme activities of human matrilysin (MMP-7) and gelatinase B/type IV collagenase (MMP-9)" *J. Biol. Chem.* 272:28823-28825 (1997)) (Stathakis, P. *et al.*, "Generation of angiostatin by reduction and proteolysis of plasmin" *J. Biol. Chem.* 272:20641-20645 (1997); Gately, S. *et al.*, "The mechanism of cancer-mediated conversion of plasminogen to that angiogenesis inhibitor angiostatin" *Proc. Natl. Acad. Sci. USA* 94:10868-10872 (1997)).

In its glycosylated state, human plasminogen protein contains an N-linked carbohydrate chain at amino acid position Asn-289 and two O-linked mucin type carbohydrate chains at amino acid positions Ser-249 and Thr-346. Since angiostatin protein is a fragment of a plasminogen protein, angiostatin protein also contains the above-described carbohydrate chains. The proteolytic digestion of plasminogen, by tPA and uPA to produce plasmin for example, is known to be modulated by the carbohydrate content of plasminogen. Further the carbohydrate chains are known to modulate binding of plasminogen to cell surface receptors. Carbohydrate is also known to play a general role in systemic half life of circulating proteins. (Mori, K. *et al.*, "The activation of type 1 and type 2 plasminogen by type I and type II tissue plasminogen activator" *J. Biol. Chem.* 270:3261-3267 (1995); Davidson, D.J. and Castellino, F.J., "The influence of the nature of the asparagine 289-linked oligosaccharide on the activation by urokinase and lysine binding properties of natural and recombinant human plasminogens" *J. Clin. Invest.* 92:249-254 (1993); Edelberg, J.

et al. "Neonatal plasminogen displays altered cell surface binding and activation kinetics" *J. Clin. Invest.* 86:107-112 (1990); Gonzales-Gronow, M. *et al.* "Further characterization of the cellular plasminogen binding site: evidence that plasminogen 2 and lipoprotein a compete for the same site" *Biochemistry*, 28:2374-2377 (1989); Jenkins, N. *et al.* "Getting the glycosylation right: implications for the biotechnology industry" *Nat. Biotechnol.* 14:975-981 (1996)).

The mechanism underlying how angiostatin and its related kringle fragments specifically inhibit endothelial cell growth remains uncharacterized. It is not yet clear whether the inhibition is mediated by a receptor that is specifically expressed in proliferating endothelial cells, or if angiostatin is internalized by endothelial cells and subsequently inhibits cell proliferation. Alternatively, angiostatin may interact with an endothelial cell adhesion receptor such as integrin $\alpha_v\beta_3$, blocking integrin-mediated angiogenesis (Brooks, P.C., *et al.* "Integrin alpha v beta 3 antagonists promote tumor regression by inducing apoptosis of angiogenic blood vessels" *Cell* 79:1157-1164 (1994)).

Although angiostatin has been identified as an angiogenesis inhibitor, what is needed in the art are kringle region fragments of plasminogen that have increased antiangiogenic activity. These improved kringle region proteins should be useful for the treatment of angiogenesis-mediated diseases, such as cancer, and for the modulation of other angiogenic processes, such as wound healing and reproduction. Due to the improved nature of the antiangiogenic kringle region proteins, these proteins should be able to be administered in smaller doses, thus lowering the cost of cancer treatment. What is also needed in the art are compositions and methods for the detection, measurement and localization of improved antiangiogenic kringle region proteins.

Summary of the Invention

Compositions and methods are provided herein that are effective for modulating angiogenesis, and inhibiting unwanted angiogenesis, especially angiogenesis related to tumor growth. In particular, the present invention provides deglycosylated kringle 1-51-3 region proteins that are fragments of plasminogen. It is a surprising discovery of the present invention that deglycosylation of kringle 1-51-3 region proteins dramatically increases the antiangiogenic activity of these proteins. It is to be understood that the deglycosylated kringle 1-51-3 region proteins described herein include fusion proteins wherein a deglycosylated kringle 1-51-3 region protein is contiguous with one or more other proteins. In a preferred embodiment, the fusion protein comprises a deglycosylated kringle 1-51-3 region protein and another antiangiogenic protein.

Also included in the present invention are nucleotides encoding the deglycosylated kringle 1-51-3 region proteins, expression vectors containing DNA sequences encoding deglycosylated kringle 1-51-3 region proteins, and cells containing one or more expression vectors containing DNA sequences encoding deglycosylated kringle 1-51-3 region proteins.

The present invention also includes antibodies specific for the deglycosylated kringle 1-51-3 region proteins, antibodies that inhibit the binding of antibodies specific for deglycosylated kringle 1-51-3 region proteins, and antibodies specific for a deglycosylated kringle 1-51-3 region protein receptor. These antibodies can be polyclonal antibodies or monoclonal antibodies. The antibodies specific for the deglycosylated kringle 1-51-3 region proteins can be used in diagnostic kits to detect the presence and quantity of deglycosylated kringle 1-51-3 region proteins which is diagnostic or prognostic for the occurrence or recurrence of cancer or other diseases mediated by angiogenesis. Antibodies specific for deglycosylated kringle

1-51-3 region proteins may also be administered to a human or animal to passively immunize the human or animal against deglycosylated kringle 1-51-3 region proteins, thereby reducing angiogenic inhibition.

5 The present invention provides methods and compositions for treating diseases and processes mediated by undesired and uncontrolled angiogenesis by increasing an *in vivo* concentration of deglycosylated kringle 1-51-3 region protein in a human or animal. *In vivo* concentrations of
10 deglycosylated kringle 1-51-3 region proteins may be increased by administering to a human or animal a composition comprising a substantially purified deglycosylated kringle 1-51-3 region protein in a dosage sufficient to inhibit angiogenesis. Additionally, *in vivo* concentrations of deglycosylated kringle 1-51-3 region proteins may be increased in a human or animal by
15 the administration of nucleotides encoding deglycosylated kringle 1-51-3 region proteins or enzymes that release deglycosylated kringle 1-51-3 region proteins from plasminogen or plasmin. The present invention is particularly
20 useful for treating or repressing the growth of tumors.

 Accordingly, it is an object of the present invention to provide a composition comprising a deglycosylated kringle 1-51-3 region protein.

25 It is another object of the present invention to provide a nucleotide composition encoding a deglycosylated kringle 1-51-3 region protein.

 It is another object of the present invention to provide compositions and methods for increasing an *in vivo* concentration of deglycosylated kringle 1-51-3 region proteins.

30 It is another object of the present invention to provide a composition comprising an antibody to a deglycosylated kringle 1-51-3 region protein that is specific for the deglycosylated kringle 1-51-3 region protein and does not recognize plasminogen.

It is an object of present invention to provide a method for detecting and quantifying the presence of an antibody specific for a deglycosylated kringle 4-51-3 region protein in a body fluid.

5 It is another object of the present invention to provide a method for the detection or prognosis of cancer.

It is another object of the present invention to provide a method of treating diseases and processes that are mediated by angiogenesis.

10 It is another object of the present invention to provide a composition for treating or repressing the growth of a cancer.

Yet another object of the invention is to provide compositions and methods useful for gene therapy for the modulation of angiogenic processes.

15 These and other objects, features and advantages of the present invention will become apparent after a review of the following detailed description of the disclosed embodiments and the appended claims.

20 **Brief Description of the Figures**

Figure 1 shows the amino acid sequence (SEQ ID NO:2) and nucleotide sequence (SEQ ID NO:1) encoding a preferred deglycosylated kringle 4-51-3 region protein.

25 Figure 2 shows the purification of glycosylated and deglycosylated kringle 4-51-3 region proteins using lys-Sepharose.

30 Figure 3 shows the purification of glycosylated and deglycosylated kringle 4-51-3 region proteins using gel filtration chromatography.

35 Figure 4 shows the inhibition of endothelial cell proliferation by glycosylated and deglycosylated kringle 4-51-3 region proteins.

Detailed Description

5 The present invention includes deglycosylated fragments of a kringle 1-51-3 region of a plasminogen protein. Also included in the present invention are methods of using the deglycosylated kringle 1-51-3 region proteins and methods and compositions for detecting the deglycosylated kringle 1-51-3 region proteins.

10 A surprising discovery of the present invention is that deglycosylated fragments of the kringle 1-51-3 region of plasminogen are dramatically more antiangiogenic than glycosylated fragments of the kringle 1-51-3 region of plasminogen. Accordingly, the present invention includes deglycosylated fragments of kringle 1-51-3 region proteins and
15 compositions comprising deglycosylated fragments of kringle 1-51-3 region proteins. In one embodiment, a composition comprises a pharmaceutically acceptable carrier and a deglycosylated fragment of a kringle 1-51-3 region of a plasminogen protein in a greater concentration than a naturally glycosylated form of the deglycosylated fragment, wherein the deglycosylated fragment lacks one or more carbohydrate
20 moieties linked to the naturally glycosylated form and wherein the deglycosylated fragment has antiangiogenic activity. In a further embodiment, the ratio of the amount of the deglycosylated fragment to the amount of the naturally glycosylated form is at least 60:40, more preferably at least 80:20, and most preferably 100:0. The ratio of the deglycosylated fragment to the naturally glycosylated form may
25 also be at least 95:5, 96:4, 97:3, 98:2 and 99:1.

30 As used herein, the term "deglycosylated" refers to a protein or peptide lacking one or more carbohydrate moieties that are linked to a naturally glycosylated form of a kringle 1-51-3 region proteins. A "naturally glycosylated form" refers to a fragment of a kringle 1-51-3 region protein, or a larger protein
35 from which the kringle 1-51-3 region fragment is derived, that

is glycosylated at amino acid positions as found in nature. For example, one naturally glycosylated form of human plasminogen is glycosylated at Ser-249, Asn-289, and Thr-346, and therefore, the naturally glycosylated form of fragments of a
5 kringle 4-51-3 region of human plasminogen are glycosylated at Ser-249, Asn-289, and/or Thr-346. It is to be understood that the naturally glycosylated forms of the kringle 4-51-3 region fragments described herein are not required to exist as a fragment in nature and can exist in nature as part of a larger
10 protein.

Therefore, "deglycosylated plasminogen" refers to a plasminogen protein of any species that lacks carbohydrate at a position that may be otherwise glycosylated in nature (i.e., that is glycosylated in a naturally glycosylated form of the
15 plasminogen protein). For example, included in the present invention is a plasminogen protein lacking a bisialylated-biantennary glycan at an amino acid position corresponding to amino acid position 289 of human plasminogen (Asn-289), an O-linked mucin type carbohydrate chain at an amino acid
20 position corresponding to amino acid position 249 of human plasminogen (Ser-249), and/or an O-linked mucin type carbohydrate chain at an amino acid position corresponding to amino acid position 346 of human plasminogen (Thr-346). Additionally, the term "deglycosylated plasminogen"
25 encompasses a plasminogen protein having zero, one or two carbohydrate chains. In one embodiment of the present invention, a deglycosylated plasminogen lacks any N-linked carbohydrate chain at amino acid corresponding to amino acid position 289 of human plasminogen. In a further embodiment,
30 a deglycosylated plasminogen lacks any carbohydrate moiety at an amino acid position corresponding to amino acid position 289 of human plasminogen.

As also used herein, the terms "deglycosylated kringle 4-51-3 region protein" and "deglycosylated fragment" refer to a
35 fragment of a kringle 4-51-3 region of a plasminogen protein of

any species that lacks carbohydrate at a position that may be otherwise glycosylated in nature (i.e., that is glycosylated in a naturally glycosylated form of the deglycosylated kringle 4-51-3 region protein). For example, included herein are
5 deglycosylated kringle 4-51-3 region proteins that lack a bisialylated-biantennary glycan at an amino acid position corresponding to amino acid position 289 of human plasminogen, an O-linked mucin type carbohydrate chain at
10 amino acid position corresponding to amino acid position 249 of human plasminogen, and/or an O-linked mucin type carbohydrate chain at amino acid position corresponding to amino acid position 346 of human plasminogen. Therefore, the term “deglycosylated kringle 4-51-3 region protein” encompasses a kringle 4-51-3 protein having zero, one or two
15 carbohydrate chains. In one embodiment of the present invention, the deglycosylated kringle 4-51-3 region protein lacks a bisialylated-biantennary glycan at amino acid position 289. In another embodiment, the carbohydrate chain at amino acid position 289 is smaller than a bisialylated-biantennary glycan. In another embodiment, the deglycosylated kringle 4-
20 51-3 region protein lacks a carbohydrate moiety at an amino acid position corresponding to amino acid position 289 of human plasminogen.

“Corresponding to”, when referring to amino acids,
25 indicates the comparison of two amino acids in the same region of different proteins, or fragments thereof, wherein the proteins are homologs, orthologs or paralogs. Homologs are defined as proteins with substantial homology, wherein “substantial homology” is defined below. Orthologs are defined as proteins
30 having non-identical amino acid sequences and similar functional characteristics, wherein the proteins are from different species, but wherein the species have a common ancestral origin. Paralogs are defined as proteins having non-identical amino acid sequences and similar functional
35 characteristics, wherein the proteins are from the same species.

5 In the present invention, an example of an amino acid corresponding to Asn-289 located in kringle 3 of human plasminogen is an amino acid located in kringle 3 of murine plasminogen that can have an N-linked carbohydrate moiety attached thereto, and preferably a bisialylated-biantennary glycan attached thereto, as found in nature. It is to be understood that the present invention includes fragments of kringle 1-51-3 regions of a plasminogen from a species other than a human.

10 In another embodiment of the present invention, the kringle 1-51-3 region protein has a modified amino acid at a position corresponding to position 289 of human plasminogen such that a carbohydrate moiety is not added to that position during post-translational modification of the protein. In a preferred embodiment, the amino acid substitution is a conservative substitution. In a further preferred embodiment, the amino acid substitution is from asparagine to glutamic acid. In yet a further preferred embodiment, the kringle 1-51-3 region protein has the amino acid sequence as shown in SEQ ID NO:2 that corresponds to an approximately kringle 1-3 region.

20 The deglycosylated kringle 1-51-3 region proteins of the present invention can be made *in vivo* or *in vitro*. For example, the deglycosylated kringle 1-51-3 region proteins can be isolated from body fluids including, but not limited to, serum, urine and ascites, or synthesized by chemical or biological methods (e.g. recombinant gene expression, chemical synthesis of oligonucleotides, and *in vitro* enzymatic catalysis of larger proteins such as plasminogen or plasmin). Recombinant techniques include gene amplification from DNA sources using the polymerase chain reaction (PCR), and gene amplification from RNA sources using reverse transcriptase/PCR. For example, a deglycosylated kringle 1-51-3 region protein can be recombinantly produced by expressing a gene encoding the protein, wherein an amino acid position that can be glycosylated in nature is substituted for an amino acid that is not

35

glycosylated. This gene may be administered to an individual using the gene therapy methods described in more detail below. The kringle 4-51-3 region proteins can also be made *in vivo* by the administration of deglycosylated plasminogen and an enzyme that cleaves deglycosylated plasminogen to an individual. In a preferred embodiment, the enzyme is an elastase enzyme such as macrophage metalloelastase. Deglycosylated kringle 4-51-3 proteins can also be made *in vivo* by the administration of an enzyme that specifically cleaves deglycosylated plasminogen.

When isolating deglycosylated kringle 4-51-3 region proteins from larger proteins such as plasminogen or plasmin, either the plasminogen or plasmin may be deglycosylated prior to enzymatic catalysis or the resulting kringle 4-51-3 region protein itself may be deglycosylated. Methods for deglycosylation of proteins are well known to those skilled in the art. However, it is a surprising discovery of the present invention that there is an increased yield of kringle 4-51-3 region proteins following enzymatic catalysis of deglycosylated plasminogen by elastase as compared to enzymatic catalysis of glycosylated plasminogen by elastase. Therefore, in a preferred embodiment of the present invention, kringle 4-51-3 region proteins are isolated by elastase digestion of deglycosylated plasminogen.

It is to be understood that "kringle 4-51-3 region" is defined herein as a region that corresponds to approximately amino acid position 1 through approximately amino acid position 541333 of human plasminogen. Thus, the deglycosylated fragments of the kringle 4-51-3 region of the present invention may contain the N-terminal sequence proceeding the kringle 1 region, kringle regions 1, 2, and 3,4, and 5, inter-kringle regions, and antiangiogenic fragments thereof, wherein the aforementioned regions are found in nature as contiguous sequences or not. Antiangiogenic fragments of

the kringle 4-51-3 region that can be deglycosylated are further disclosed in U.S. Patent Nos. 5,639,725 and 5,837,682.

5 It is also to be understood that the present invention is contemplated to include any deglycosylated kringle 4-51-3 region protein derivatives. A kringle 4-51-3 region protein derivative includes a protein having the amino acid sequence of a kringle 4-51-3 region of a plasminogen protein. A kringle 4-51-3 region protein derivative also includes a peptide having a sequence corresponding to an antiangiogenic fragment a kringle
10 4-51-3 region. An "antiangiogenic fragment" is defined to be a peptide whose amino acid sequence corresponds to a subsequence of a kringle 4-51-3 region, referred to as an "antiangiogenic subsequence". A "subsequence" is a sequence of contiguous amino acids found within a larger sequence. A
15 subsequence is generally composed of approximately at least 70%, more preferably 80%, and most preferably 90% of the larger sequence.

A kringle 4-51-3 region protein derivative also includes a protein or peptide having a modified sequence in which one or
20 more amino acids in the original sequence or subsequence have been substituted with a naturally occurring amino acid residue or amino acid residue analog (also referred to as modified amino acid). Suitable kringle 4-51-3 region protein derivatives have modified sequences which are substantially homologous to
25 the amino acid sequence of a kringle 4-51-3 region protein or to an antiangiogenic subsequence of a kringle 4-51-3 region protein.

An "amino acid residue" is a moiety found within a protein or peptide and is represented by -NH-CHR-CO-,
30 wherein R is the side chain of a naturally occurring amino acid. When referring to a moiety found within a peptide, the terms "amino acid residue" and "amino acid" are used interchangeably. An "amino acid residue analog" includes D or L configurations having the following formula: -NH-CHR-CO-,
35 wherein R is an aliphatic group, a substituted aliphatic

aromatic group, a benzyl group, a substituted benzyl group, an aromatic group or a substituted aromatic group and wherein R does not correspond to the side chain of a naturally occurring amino acid.

5 Suitable substitutions for amino acid residues in the sequence of the deglycosylated kringle 1-51-3 region proteins described herein include conservative substitutions that result in angiogenic deglycosylated kringle 1-51-3 region protein derivatives. A conservative substitution is a substitution in
10 which the substituting amino acid (naturally occurring or modified) is structurally related to the amino acid being substituted. "Structurally related" amino acids are approximately the same size and have the same or similar functional groups in the side chains.

15 Provided below are groups of naturally occurring and modified amino acids in which each amino acid in a group has similar electronic and steric properties. Thus, a conservative substitution can be made by substituting an amino acid with another amino acid from the same group. It is to be understood
20 that these groups are non-limiting and that additional modified amino acids could be included in each group.

 Group I includes leucine, isoleucine, valine, methionine and modified amino acids having the following side chains: ethyl, *n*-propyl *n*-butyl. Preferably, Group I includes leucine,
25 isoleucine, valine and methionine.

 Group II includes glycine, alanine, valine and a modified amino acid having an ethyl side chain. Preferably, Group II includes glycine and alanine.

30 Group III includes phenylalanine, phenylglycine, tyrosine, tryptophan, cyclohexylmethyl, and modified amino residues having substituted benzyl or phenyl side chains. Preferred substituents include one or more of the following: halogen, methyl, ethyl, nitro, -NH₂, methoxy, ethoxy and -CN. Preferably, Group III includes phenylalanine, tyrosine and
35 tryptophan.

Group IV includes glutamic acid, aspartic acid, a substituted or unsubstituted aliphatic, aromatic or benzylic ester of glutamic or aspartic acid (e.g., methyl, ethyl, *n*-propyl *iso*-propyl, cyclohexyl, benzyl or substituted benzyl), glutamine, asparagine, -CO-NH-alkylated glutamine or asparagine (e.g., methyl, ethyl, *n*-propyl and *iso*-propyl) and modified amino acids having the side chain $-(CH_2)_3-COOH$, an ester thereof (substituted or unsubstituted aliphatic, aromatic or benzylic ester), an amide thereof and a substituted or unsubstituted N-alkylated amide thereof. Preferably, Group IV includes glutamic acid, aspartic acid, methyl aspartate, ethyl aspartate, benzyl aspartate and methyl glutamate, ethyl glutamate and benzyl glutamate, glutamine and asparagine.

Group V includes histidine, lysine, ornithine, arginine, N-nitroarginine, β -cycloarginine, γ -hydroxyarginine, N-amidinocitruline and 2-amino-4-guanidinobutanoic acid, homologs of lysine, homologs of arginine and homologs of ornithine. Preferably, Group V includes histidine, lysine, arginine and ornithine. A homolog of an amino acid includes from 1 to about 3 additional or subtracted methylene units in the side chain.

Group VI includes serine, threonine, cysteine and modified amino acids having C1-C5 straight or branched alkyl side chains substituted with -OH or -SH, for example, -CH₂CH₂OH, -CH₂CH₂CH₂OH or -CH₂CH₂OHCH₃. Preferably, Group VI includes serine, cysteine or threonine.

In another aspect of the present invention, suitable substitutions for amino acid residues in the amino acid sequences described herein include "severe substitutions" that result in kringle 4-51-3 region protein derivatives that are antiangiogenic. Severe substitutions that result in antiangiogenic kringle 4-51-3 region protein derivatives are much more likely to be possible in positions that are not highly conserved than at positions that are highly conserved. In the present invention, severe substitutions are much more likely to

be possible in the inter-kringle regions and the N-terminal sequence proceeding kringle 1. A "severe substitution" is a substitution in which the substituting amino acid (naturally occurring or modified) has significantly different size and/or electronic properties compared with the amino acid being substituted. For example, the side chain of the substituting amino acid can be significantly larger (or smaller) than the side chain of the amino acid being substituted and/or can have functional groups with significantly different electronic properties than the amino acid being substituted.

Examples of severe substitutions of this type include the substitution of phenylalanine or cyclohexylmethyl glycine for alanine, isoleucine for glycine, a D amino acid for the corresponding L amino acid or $-\text{NH}-\text{CH}[(\text{-CH}_2)_5\text{-COOH}]-\text{CO}-$ for aspartic acid. Alternatively, a functional group may be added to the side chain, deleted from the side chain or exchanged with another functional group. Examples of severe substitutions of this type include adding an amine or hydroxyl, carboxylic acid to the aliphatic side chain of valine, leucine or isoleucine, exchanging the carboxylic acid in the side chain of aspartic acid or glutamic acid with an amine or deleting the amine group in the side chain of lysine or ornithine. In yet another alternative, the side chain of the substituting amino acid can have significantly different steric and electronic properties than the functional group of the amino acid being substituted. Examples of such modifications include tryptophan for glycine, lysine for aspartic acid and $-(\text{CH}_2)_4\text{COOH}$ for the side chain of serine. These examples are not meant to be limiting.

"Substantial homology" exists between two amino acid sequences when a sufficient number of amino acid residues at corresponding positions of each amino acid sequence are either identical or structurally related such that a protein or peptide having the first amino acid sequence and a protein or peptide having the second amino acid sequence exhibit similar biological activities. Generally, there is substantial sequence

homology among the amino acid sequences when at least 30%, more preferably at least 40%, and most preferably at least 50%, of the amino acids in the first amino acid sequence are identical to or structurally related to the second amino acid sequence.

5 Homology is often measured using sequence analysis software, e.g., BLASTIN or BLASTP (available at the National Institutes of Health (NIH) World Wide Web Server~~<http://www.ncbi.nlm.nih.gov/BLAST>~~). The default parameters for comparing the two sequences (e.g., "Blast"-ing

10 two sequences against each other) by BLASTIN (for nucleotide sequences) are reward for match = 1, penalty for mismatch = -2, open gap = 5, and extension gap = 2. When using BLASTP for protein sequences, the default parameters are reward for match = 0, penalty for mismatch = 0, open gap = 11, and extension gap = 1.

15

With regard to the present invention, the kringle region motif of kringles 1-3 of a plasminogen protein is also known to exist in other biologically active proteins. These proteins include, but are not limited to, prothrombin, hepatocyte growth

20 factor, scatter factor and macrophage stimulating protein. (Yoshimura, T, *et al.*, "Cloning, sequencing, and expression of human macrophage stimulating protein (MSP, MST1) confirms MSP as a member of the family of kringle proteins and locates the MSP gene on Chromosome 3" *J. Biol. Chem.*, 268:15461-15468, (1993)). It is contemplated that any kringle ~~4-5~~1-3

25 region protein or peptide having a three-dimensional kringle-like conformation or cysteine motif that has antiangiogenic activity *in vivo*, is part of the present invention.

It is to be understood that as used herein, the term

30 "isolated" refers to a composition which is substantially or essentially free from at least some of the components that normally accompany it in its native state. Thus, the deglycosylated kringle ~~4-5~~1-3 region proteins of this invention do not contain some of the materials normally associated with

35 their *in situ* environment. Typically, the isolated,

deglycosylated kringle region 1-5 proteins of the invention are at least about 80% pure, usually at least about 90% pure, and preferably at least about 95% pure as measured by band intensity on a silver stained gel. It is to be understood that the term “isolated” does not exclude fusion proteins comprising the deglycosylated kringle 1-51-3 region proteins from the present invention. The present invention contemplates fusion proteins, or chimeric proteins, comprising the kringle 1-51-3 region proteins described herein and other proteins. In a preferred embodiment the fusion proteins comprise deglycosylated kringle 1-51-3 region proteins and other antiangiogenic proteins such as endostatin proteins, wherein the endostatin proteins are defined as antiangiogenic fragments of the C-terminal non-collagenous region of a collagen protein.

The deglycosylated kringle 1-51-3 region proteins described herein are useful for treating diseases or processes that are mediated by, or involve, angiogenesis. In particular, the deglycosylated kringle 1-51-3 region proteins are useful for the treatment of angiogenesis-mediated cancers. The deglycosylated kringle 1-51-3 region proteins are also useful for the generation of antibodies specific for the deglycosylated kringle 1-51-3 region proteins, and for the identification and isolation of receptors for and chemical mimetics of deglycosylated kringle 1-51-3 region proteins.

In addition to deglycosylated kringle 1-51-3 region proteins, the present invention encompasses compositions comprising nucleotide sequences encoding the kringle 1-51-3 region proteins described herein. In one embodiment of the present invention, the nucleotide sequence encodes a kringle 1-51-3 region protein containing a modified amino acid at a position corresponding to amino acid 289 of human plasminogen. In a preferred embodiment, the nucleotide sequence is as shown in SEQ ID NO:2. NO:1. The present invention also includes a vector containing a DNA sequence encoding a kringle 1-51-3 region protein, wherein the vector is

capable of expressing kringle 4-51-3 region proteins when present in a cell. The cell may contain one vector or multiple vectors. The nucleotide sequences described herein are useful for recombinantly producing the deglycosylated kringle 4-51-3 region proteins of the present invention and for treatment of angiogenesis associated diseases and conditions via gene therapy.

Still further, the deglycosylated kringle 4-51-3 region proteins can be used to generate antibodies to the deglycosylated kringle 4-51-3 region proteins and their receptors. In particular, an antibody can be generated that specifically binds to a deglycosylated kringle 4-51-3 region but does not bind to glycosylated plasminogen or a glycosylated kringle 4-51-3 region protein. The terms "antibody" and "antibodies" as used herein include monoclonal, polyclonal, chimeric, single chain, bispecific, simianized, and humanized antibodies as well as Fab fragments, including the products of an Fab immunoglobulin expression library.

To enhance the potential for high specificity in the development of antisera (or agonists and antagonists) to deglycosylated kringle 4-51-3 region proteins, protein sequences can be compared to known sequences using protein sequence databases such as GenBank, Brookhaven Protein, SWISS-PROT, and PIR to determine potential sequence homologies. This information facilitates elimination of sequences that exhibit a high degree of sequence homology to other molecules. These antibodies that specifically bind to the deglycosylated kringle 4-51-3 region proteins or their receptors, can be used in diagnostic methods and kits that are well known to those of ordinary skill in the art to detect or quantify the deglycosylated kringle 4-51-3 region proteins or their receptors in a body fluid or tissue. Results from these tests can be used to diagnose or predict the occurrence or recurrence of a cancer or other angiogenic-mediated disease.

The phrases "specifically binds to" or "specific for" when referring to an antibody refers to a binding reaction which is determinative of the presence of the peptide in the presence of a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies bind preferentially to a particular peptide and do not bind in a significant amount to other proteins present in the sample. Specific binding to a peptide under such conditions requires an antibody that is selected for its specificity for a particular protein. A variety of immunoassay formats may be used to select antibodies specifically immunoreactive with a particular protein. For example, solid-phase ELISA immunoassays are routinely used to select monoclonal antibodies specifically immunoreactive with a protein. See, Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring Harbor Publications, New York, for a description of immunoassay formats and conditions that can be used to determine specific immunoreactivity.

When labeled with a detectable biomolecule or chemical, the deglycosylated kringle 1-51-3 region proteins, nucleic acids and antibodies described above are useful for purposes such as *in vivo* and *in vitro* diagnostics and laboratory research using the methods and assays described below. Various types of labels and methods of conjugating the labels to the polypeptides and antibodies are well known to those skilled in the art. Examples of labels include, but are not limited to, radiolabels, bioluminescent labels, fluorogens and chromogens. Still further, the present invention encompasses deglycosylated kringle 1-51-3 region proteins, deglycosylated kringle 1-51-3 region protein antisera, deglycosylated kringle 1-51-3 region protein receptor agonists or deglycosylated kringle 1-51-3 region protein receptor antagonists that are combined with pharmaceutically acceptable excipients, and optionally sustained-release compounds or compositions, such as

biodegradable polymers, to form therapeutic compositions for use in the methods of treatment described below.

5 The methods of the present invention include methods of making, detecting and measuring the deglycosylated kringle 4-51-3 region proteins of the present invention as well as methods of treating angiogenesis associated conditions and diseases using the deglycosylated kringle 4-51-3 region proteins of the present invention. Deglycosylated kringle 4-51-3 region proteins can be produced synthetically by chemical reaction or
10 by recombinant techniques in conjunction with expression systems. These deglycosylated kringle 4-51-3 region proteins can be animal or human in origin. In one embodiment, the deglycosylated kringle 4-51-3 region protein is produced recombinantly and has the amino acid sequence of SEQ ID
15 NO:2.

Deglycosylated kringle 4-51-3 region proteins may also be produced *in vitro* or *in vivo* by enzymatically cleaving plasminogen or plasmin to generate proteins having antiangiogenic activity or by using compounds that mimic the
20 action of endogenous enzymes that cleave deglycosylated plasminogen into deglycosylated kringle 4-51-3 region proteins. Deglycosylated kringle 4-51-3 region protein production may also be modulated by compounds that affect the activity of plasminogen cleaving enzymes. In particular, it is a surprising
25 discovery of the present invention that glycosylation of plasminogen reduces the yield of kringle 4-51-3 region proteins upon cleavage of plasminogen with elastase. Therefore, in one embodiment of the present invention, the method of making deglycosylated kringle 4-51-3 region proteins comprises
30 cleaving deglycosylated plasminogen with elastase. Given the role of elastase reported here in the generation of kringle 4-51-3 region proteins, it is likely that the N-linked carbohydrate on kringle 3 will also modulate the reaction mechanism of other proteinases potentially responsible for kringle 4-51-3 region
35 protein generation.

5 The present invention also includes methods for the
detection of deglycosylated kringle 4-51-3 region proteins in
body fluids and tissues for the purpose of determining the
efficacy of treatment and for the prognosis and diagnosis of
angiogenesis associated diseases such as cancer. In particular,
provided herein are methods of using antibodies that are
specific for deglycosylated kringle 4-51-3 region proteins over
circulating plasminogen. These methods provide a means to
detect and distinguish deglycosylated kringle 4-51-3 region
10 proteins from glycosylated kringle 4-51-3 region proteins and
glycosylated plasminogen. The present invention also includes
the detection of deglycosylated kringle 4-51-3 region protein
binding sites and receptors in cells and tissues.

15 Kits for detection and measurement of deglycosylated
kringle 4-51-3 region proteins, and the receptors therefor, are
contemplated as part of the present invention. Antisera that
possess the highest titer and specificity and can detect
deglycosylated kringle 4-51-3 region proteins in extracts of
plasma, urine, tissues, and in cell culture media are further
20 examined to establish easy to use kits for rapid, reliable,
sensitive, and specific measurement and localization of
deglycosylated kringle 4-51-3 region proteins. These assay kits
include but are not limited to the following techniques;
competitive and non-competitive assays, radioimmunoassay,
25 bioluminescence and chemiluminescence assays, fluorometric
assays, sandwich assays, immunoradiometric assays, dot blots,
enzyme linked assays including ELISA, antibody coated strips
or dipsticks for rapid monitoring of urine or blood, and
immunocytochemistry. For each kit the range, sensitivity,
30 precision, reliability, specificity and reproducibility of the assay
are established. Intra-assay and inter-assay variation is
established at 20%, 50% and 80% points on the standard curves
of displacement or activity.

35 The present invention also includes methods of treating
or preventing angiogenic diseases and processes including, but

not limited to, arthritis and tumors by stimulating the production of deglycosylated kringle 4-51-3 region proteins, and/or by administering substantially purified deglycosylated kringle 4-51-3 region proteins, nucleotides encoding deglycosylated kringle 4-51-3 region proteins, or deglycosylated kringle 4-51-3 region protein agonists or antagonists, and/or deglycosylated kringle 4-51-3 region protein antisera or antisera directed against deglycosylated kringle 4-51-3 region protein antisera to a patient. These angiostatin deglycosylated kringle 4-51-3 region proteins, deglycosylated kringle 4-51-3 region protein antisera, deglycosylated kringle 4-51-3 region protein receptor agonists or antagonists, or combinations thereof, are combined with pharmaceutically acceptable excipients, and optionally a sustained-release matrix, such as biodegradable polymers, to form therapeutic compositions. Additional treatment methods include administration of deglycosylated kringle 1-53 proteins, deglycosylated kringle 4-51-3 region protein analogs, deglycosylated kringle 4-51-3 region protein antisera, or deglycosylated kringle 4-51-3 region protein receptor agonists and antagonists linked to cytotoxic agents.

In one aspect of the present invention, a method of inhibiting angiogenesis in an individual comprises, increasing in the individual an *in vivo* concentration of a deglycosylated fragment of a kringle 4-51-3 region of a plasminogen protein relative to the *in vivo* concentrations of a glycosylated fragment of a kringle 4-51-3 region of a plasminogen protein and wherein the deglycosylated fragment of the kringle 4-51-3 region protein has antiangiogenic activity *in vivo*. The *in vivo* concentration of the deglycosylated fragment can be increased in any bodily fluid or tissue, but is preferably increased in the serum. In a further embodiment, a deglycosylated kringle 4-51-3 region protein is administered in a treatment effective amount to an individual in need of such treatment. In another embodiment, a gene encoding a deglycosylated fragment of a kringle 4-51-3 region of a plasminogen is administered to an individual using

the gene therapy methods discussed in more detail below. Preferably the gene contains an amino acid substitution that prevents glycosylation during post-translational modification of the fragment of the kringle 4-51-3 region, and more preferably the substitution is at an amino acid corresponding to amino acid Asn-289 of human plasminogen.

A deglycosylated kringle 4-51-3 region protein is effective in treating diseases or processes that are mediated by, or involve, angiogenesis. The present invention includes the method of treating an angiogenesis mediated disease with an effective amount of a deglycosylated kringle 4-51-3 region protein, or combinations of deglycosylated kringle 4-51-3 region proteins that collectively possess antiangiogenic activity, or deglycosylated kringle 4-51-3 region protein agonists and antagonists. The angiogenesis mediated diseases include, but are not limited to, solid tumors; blood born tumors such as leukemias; tumor metastasis; benign tumors, for example hemangiomas, acoustic neuromas, neurofibromas, trachomas, and pyogenic granulomas; rheumatoid arthritis; psoriasis; ocular angiogenic diseases, for example, diabetic retinopathy, retinopathy of prematurity, macular degeneration, corneal graft rejection, neovascular glaucoma, retrolental fibroplasia, rubeosis; Osler-Webber Syndrome; myocardial angiogenesis; plaque neovascularization; telangiectasia; hemophiliac joints; angiofibroma; and wound granulation. Deglycosylated kringle 4-51-3 region protein is useful in the treatment of disease of excessive or abnormal stimulation of endothelial cells. These diseases include, but are not limited to, intestinal adhesions, Crohn's disease, atherosclerosis, scleroderma, and hypertrophic scars, i.e., keloids. Deglycosylated kringle 4-51-3 region proteins can be used as a birth control agent by preventing vascularization required for embryo implantation. Deglycosylated kringle 4-51-3 region proteins are also useful in the treatment of diseases that have angiogenesis as a pathologic

consequence such as cat scratch disease (*Rochela minalia quintosa*) and ulcers (*Helicobacter pylori*).

5 Deglycosylated kringle 4-51-3 region proteins may be used in combination with other compositions and procedures for the treatment of diseases. For example, a tumor may be treated conventionally with surgery, radiation or chemotherapy combined with deglycosylated kringle 4-51-3 region proteins and then deglycosylated kringle 4-51-3 region proteins may be subsequently administered to the patient to extend the dormancy of micrometastases and to stabilize and inhibit the growth of any residual primary tumor.

10 In addition to methods of administering deglycosylated kringle 4-51-3 region proteins, included in the present invention are methods of passive antibody therapy. In one embodiment, deglycosylated kringle 4-51-3 region protein antibodies block the action of excess endogenous deglycosylated kringle 4-51-3 region proteins. Specifically blocking the action of excess endogenous deglycosylated kringle 4-51-3 region proteins (versus endogenous glycosylated kringle 4-51-3 region proteins) is important considering the novel teaching herein that deglycosylated kringle 4-51-3 region proteins are more antiangiogenic than glycosylated kringle 4-51-3 region proteins. This treatment provides an improved method of modulating angiogenic-dependent processes such as reproduction, development, and wound healing and tissue repair and treating abnormal ovulation, menstruation and placentation, and vasculogenesis. This method also provides a useful tool to examine the effects of deglycosylated kringle 4-51-3 region protein removal on metastatic processes.

25 In another embodiment, antisera directed to the Fab regions of deglycosylated kringle 4-51-3 region protein antibodies can be administered to block the ability of endogenous deglycosylated kringle 4-51-3 region protein antisera to bind deglycosylated kringle 4-51-3 region proteins. The net effect of this treatment is to facilitate the ability of

endogenous circulating deglycosylated kringle 1-51-3 region protein to reach target cells, thereby decreasing the spread of metastases.

5 The deglycosylated kringle 1-51-3 region proteins and antibodies thereto described above can be provided as isolated and substantially purified proteins and protein fragments in pharmaceutically acceptable formulations using formulation methods known to those of ordinary skill in the art. The
10 deglycosylated kringle 1-51-3 region proteins and antibodies thereto described above may be a solid, liquid or aerosol. Examples of solid therapeutic compositions include pills, creams, and implantable dosage units. The pills may be administered orally and the therapeutic creams may be administered topically. The implantable dosage units may be
15 administered locally, for example at a tumor site, or may be implanted for systemic release of the therapeutic angiogenesis-modulating composition, for example subcutaneously. Examples of liquid compositions include formulations adapted for injection subcutaneously, intravenously, intraarterially, and
20 formulations for topical and intraocular administration. Examples of aerosol formulations include inhaler formulations for administration to the lungs. In general however, the formulations may be administered by any route, including but not limited to, the topical, transdermal, intraperitoneal,
25 intracranial, intracerebroventricular, intracerebral, intravaginal, intrauterine, oral, rectal or parenteral (e.g., intravenous, intraspinal, subcutaneous or intramuscular) route.

 In addition, the deglycosylated kringle 1-51-3 region proteins and antibodies thereto may be incorporated into
30 biodegradable polymers allowing for sustained release of the compound, the polymers being implanted in the vicinity of where drug delivery is desired, for example, at the site of a tumor or implanted so that the deglycosylated kringle 1-51-3 region protein or antibody is slowly released systemically. The
35 biodegradable polymers and their use are described, for

example, in detail in Brem *et al.*, "Interstitial chemotherapy with drug polymer implants for the treatment of recurrent gliomas" *J. Neurosurg.* 74:441-446 (1991). Osmotic minipumps may also be used to provide controlled delivery of high concentrations of deglycosylated kringle 4-51-3 region proteins and antibodies thereto through cannulae to the site of interest, such as directly into a metastatic growth or into the vascular supply to that tumor.

The dosage of the deglycosylated kringle 4-51-3 region proteins and antibodies of the present invention will depend on the disease state or condition being treated and other clinical factors such as weight and condition of the human or animal and the route of administration of the compound. For treating humans or animals, between approximately 0.5 mg/kilogram per day to 500 mg/kilogram per day of the deglycosylated kringle 4-51-3 region protein can be administered. Depending upon the half-life of the deglycosylated kringle 4-51-3 region protein in the particular animal or human, it can be administered between several times per day to once a week. It is to be understood that the present invention has application for both human and veterinary use. The methods of the present invention contemplate single as well as multiple administrations, given either simultaneously or over an extended period of time.

The deglycosylated kringle 4-51-3 region protein formulations may conveniently be presented in unit dosage form and may be prepared by conventional pharmaceutical techniques. Such techniques include the step of bringing into association the active ingredient and the pharmaceutical carrier(s) or excipient(s). Suitable pharmaceutical carriers and excipients are known to those skilled in the art, however, an example of a suitable pharmaceutical excipient is water. In general, the formulations are prepared by uniformly and intimately bringing into association the active ingredient with

liquid carriers or finely divided solid carriers or both, and then, if necessary, shaping the product.

5 Formulations suitable for parenteral administration include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents. The formulations may be presented in unit-
10 dose or multi-dose containers, for example, sealed ampules or vials, and may be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid carrier, for example, water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be
15 prepared from sterile powders, granules and tablets of the kind previously described.

Preferred unit dosage formulations are those containing a daily dose or unit, daily sub-dose, or an appropriate fraction thereof, of the administered ingredient. It should be understood
20 that in addition to the ingredients, particularly mentioned above, the formulations of the present invention may include other agents conventional in the art having regard to the type of formulation in question. Optionally, cytotoxic agents may be incorporated or otherwise combined with deglycosylated
25 kringle 4-51-3 region proteins, or biologically functional protein fragments thereof, to provide dual therapy to the patient.

The present invention also encompasses gene therapy whereby the gene encoding a deglycosylated kringle 4-51-3 region protein is regulated in a patient. Various methods of
30 transferring or delivering DNA to cells for expression of the gene product, otherwise referred to as gene therapy, are disclosed in "Gene Transfer into Mammalian Somatic Cells *in vivo*", N. Yang, *Crit. Rev. Biotechn.* 12(4):335-356 (1992). Gene therapy encompasses incorporation of DNA sequences
35 into somatic cells or germ line cells for use in either *ex vivo* or

in vivo therapy. Gene therapy functions to replace genes, augment normal or abnormal gene function, and to combat infectious diseases and other pathologies.

5 Strategies for treating medical problems with gene therapy include therapeutic strategies such as replacing a defective gene with a functional gene or adding a functional gene to augment a slightly functional gene. The genes inserted may treat a disease or condition or make the tissue or organ more susceptible to a treatment regimen. With regard to the
10 present invention, a gene for a deglycosylated kringle 4-51-3 region protein may be placed in a subset of cells, thus preventing the occurrence of angiogenesis in the transformed cells.

Many protocols for the transfer of deglycosylated kringle
15 4-51-3 region protein DNA or deglycosylated kringle 4-51-3 region protein regulatory sequences are envisioned in this invention. Transfection of promoter sequences, other than one normally found specifically associated with a deglycosylated kringle 4-51-3 region protein, or other sequences that would
20 increase production of deglycosylated kringle 4-51-3 region proteins are envisioned as methods of gene therapy. Such "genetic switches" could be used to activate a deglycosylated kringle 4-51-3 region protein (or the deglycosylated kringle 4-51-3 region protein receptor) in cells not normally expressing
25 deglycosylated kringle 4-51-3 region protein (or the deglycosylated kringle 4-51-3 region protein receptor).

Gene transfer methods for gene therapy fall into three broad categories: (1) chemical (lipid-based carriers, or other non-viral vectors), (2) biological (virus-derived vector and
30 receptor uptake), and (3) physical (electroporation, direct gene transfer and particle bombardment). Gene therapy methodologies can also be described by delivery site. Fundamental ways to deliver genes include *ex vivo* gene transfer, *in vitro* gene transfer, and *in vivo* gene transfer. In *ex vivo*
35 gene transfer, cells are taken from the patient and grown in

cell culture. The DNA is transfected into the cells, the transfected cells are expanded in number and then re-implanted in the patient. The present invention encompasses the removal of endothelial cells from a patient, transfection of DNA
5 encoding a deglycosylated kringle ~~1-51~~1-3 region protein, or regulatory sequence thereof, and re-introduction of the transfected endothelial cells into the patient. In *in vitro* gene transfer, transformed cells, such as endothelial cells, growing in culture are introduced into the patient. The transformed cells
10 are not taken from the patient who will receive the gene therapy. *In vivo* gene transfer involves introducing the DNA into the cells of the patient when the cells are within the patient. Methods include using a noninfectious virus to introduce a gene into a patient or injecting naked DNA into a site in the patient
15 whereby DNA is taken up by a percentage of cells in which the gene product protein is expressed. In the present invention, DNA encoding a deglycosylated kringle ~~1-51~~1-3 region protein can be introduced into the endothelial cells lining the blood vessels, thereby inhibiting angiogenesis. In a preferred
20 embodiment, the DNA encoding a deglycosylated kringle ~~1-51~~1-3 region protein is introduced into endothelial cells lining the blood vessels in close proximity to a tumor.

Chemical methods of gene therapy may involve a lipid based compound, not necessarily a liposome, used to ferry the
25 DNA across the cell membrane. Lipofectins or cytofectins, lipid-based positive ions that bind to negatively charged DNA, make a complex that can cross the cell membrane and provide the DNA into the interior of the cell. Liposome/DNA complexes may be directly injected intravenously into the
30 patient. It is believed that the liposome/DNA complexes are concentrated in the liver where they deliver the DNA to macrophages and Kupffer cells. These cells are long lived and thus provide long term expression of the delivered DNA. Additionally, vectors or the "naked" DNA of the gene may be
35 directly injected into the desired organ, tissue or tumor for

targeted delivery of the therapeutic DNA. Other DNA carrier systems include the asialoglycoprotein/polylysine conjugate system for carrying DNA to hepatocytes for *in vivo* gene transfer and DNA coupled to nuclear proteins in specifically engineered vesicle complexes that are carried directly into the nucleus.

Biological methods used in gene therapy techniques may involve receptor-based endocytosis, or receptor-based phagocytosis, which involve binding a specific ligand to a cell surface receptor and enveloping and transporting the ligand across the cell membrane. Specifically, a ligand/gene complex is created and injected into the blood stream. Target cells having a receptor for the ligand will specifically bind the ligand and transport the ligand-DNA complex into the cell. Additional biological methods employ viral vectors to insert genes into cells. For example, altered retrovirus vectors have been used in *ex vivo* methods to introduce genes into peripheral and tumor-infiltrating lymphocytes, hepatocytes, epidermal cells, myocytes, and other somatic cells. These altered cells are then introduced into the patient.

Viral vectors have also been used to insert genes into cells using *in vivo* protocols. To accomplish tissue-specific expression of foreign genes, cis-acting regulatory elements or promoters that are known to be tissue specific can be used. Alternatively, tissue-specific expression can be achieved using *in situ* delivery of DNA or viral vectors to specific anatomical sites *in vivo*. For example, gene transfer to blood vessels *in vivo* has been achieved by implanting *in vitro* transduced endothelial cells in chosen sites on arterial walls. Surrounding cells were infected by the virus and therefore also expressed the gene product. A viral vector can be delivered directly to the *in vivo* site, by a catheter for example, thus allowing only certain areas to be infected by the virus. *In vivo* gene transfer using retrovirus vectors has also been demonstrated in mammary

tissue and hepatic tissue by injection of the altered virus into blood vessels leading to the organs.

5 Viral vectors that have been used for gene therapy protocols include, but are not limited to, retroviruses such as murine leukemia retroviruses, RNA viruses such as poliovirus or Sindbis virus, adenovirus, adeno-associated virus, herpes
10 viruses, SV40, vaccinia and other DNA viruses. Replication-defective murine retroviral vectors are the most widely utilized gene transfer vectors. Fundamental advantages of retroviral
15 vectors for gene transfer include efficient infection and gene expression in most cell types, precise single copy vector integration into target cell chromosomal DNA, and ease of manipulation of the retroviral genome. The adenovirus is capable of transducing novel genetic sequences into target cells
20 *in vivo*. Adenoviral-based vectors express gene product proteins at high levels and have high efficiencies of infectivity, even with low titers of virus. Additionally, the virus is fully infective as a cell free virion so injection of expression cell lines is not necessary. Another potential advantage to adenoviral
25 vectors is the ability to achieve long term expression of heterologous genes *in vivo*.

 Mechanical methods of DNA delivery include direct injection of DNA, such as microinjection of DNA into germ or somatic cells, pneumatically delivered DNA-coated particles,
30 such as the gold particles used in a "gene gun," inorganic chemical approaches such as calcium phosphate transfection and electroporation. It has been found that injecting plasmid DNA into muscle cells yields high percentage of the cells that are transfected and have sustained expression of marker genes.
35 The DNA of the plasmid may or may not integrate into the genome of the cells. Non-integration of the transfected DNA would allow the transfection and expression of gene product proteins in terminally differentiated, non-proliferative tissues for a prolonged period of time without fear of mutational insertions, deletions, or alterations in the cellular or

mitochondrial genome. Long-term, but not necessarily permanent, transfer of therapeutic genes into specific cells may provide treatments for genetic diseases or for prophylactic use. The DNA could be re-injected periodically to maintain the gene product level without mutations occurring in the genomes of the recipient cells. Non-integration of exogenous DNAs may allow for the presence of several different exogenous DNA constructs within one cell with all of the constructs expressing various gene products.

Both particle-mediated gene transfer methods and electroporation can be used in *in vitro* systems, or with *ex vivo* or *in vivo* techniques to introduce DNA into cells, tissues or organs. With regard to particle-mediated gene transfer, a particle bombardment device, or "gene gun," is used that generates a motive force to accelerate DNA-coated high density particles (such as gold or tungsten). These particles penetrate the target organs, tissues or cells. Electroporation mediated gene transfer comprises the use of a brief electric impulse with a given field strength that is used to increase the permeability of a membrane in such a way that DNA molecules can penetrate into the cells.

The gene therapy protocol for transfecting DNA encoding deglycosylated kringle 4-51-3 region proteins into a patient may either be through integration of the deglycosylated kringle 4-51-3 region protein DNA into the genome of the cells, into minichromosomes or as a separate replicating or non-replicating DNA construct in the cytoplasm or nucleoplasm of the cell. Deglycosylated kringle 4-51-3 region protein expression may continue for a long-period of time or the DNA may be re-injected periodically to maintain a desired level of the deglycosylated kringle 4-51-3 region protein in serum or in a cell, tissue or organ.

This invention is further illustrated by the following examples, which are not to be construed in any way as imposing limitations upon the scope thereof. It is to be clearly understood

that resort may be had to various other embodiments, modifications, and equivalents thereof which, after reading the description herein, may suggest themselves to those skilled in the art without departing from the spirit of the present invention and/or the scope of the appended claims.

5

Example 1

Isolation of glycosylated and deglycosylated kringle 4-51-3 region proteins

10 Human plasminogen was purified using affinity chromatography as described in Brockway, W.J. and Castellino, F.J., "Measurement and the binding of antifibrinolytic amino acids to various plasminogens" *Arch. Biochem. Biophys.* 151:194-199 (1972). One liter of human plasma (Children's Hospital blood bank) was applied to a 200mL lys-Sepharose 4b (Pharmacia) column at a flow rate of 2-3 ml/minute. The column, previously equilibrated with 50mM Tris pH 7.4 (Sigma) was then washed with 500mL 50mM tris/1M NaCl pH 7.4. Bound plasminogen was eluted with 200mL 50m M tris/200mM ϵ -aminocaproic acid (ϵ ACA). The purity of this material was greater than 95% as assessed by SDS-PAGE. Human plasminogen was further fractionated into glycosylated plasminogen (plasminogen 1) and deglycosylated plasminogen (plasminogen 2) using lectin affinity chromatography. Human plasminogen (10mg) was applied to a 5mL conA HiTrap column (Pharmacia), equilibrated with 50mM Tris pH 7.4/1mM $MgCl_2$ /1mM $CaCl_2$. Plasminogen 2, lacking an N-linked carbohydrate, does not bind to this column. After washing the column with five column volumes of equilibration buffer, plasminogen 1 was eluted using a solution of 50mM Tris pH 7.4/1mM $MgCl_2$ /1mM $CaCl_2$. Protein concentration was determined spectrophotometrically as a wavelength of 280nm using an $A^{0.1\%/1cm}$ of 1.6 (computed from the protparam tool available at <http://www.expasy.ch/>).

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Plasminogen 1 and 2 were dialyzed against 20mM Tris pH 7.6. Equal amounts of these glycoforms (as determined by A280nm) were then digested with porcine pancreatic elastase (PPE) as described in O'Reilly, M.S. *et al.*, "Angiostatin induces and sustains dormancy of human primary tumors in mice" *Nature Medicine* 2:689-692 (1996). Essentially, plasminogen was digested at 37°C with 0.8 units of PPE (Calbiochem) per milligram of plasminogen. After five hours incubation, the reaction was quenched by applying the reaction mixture to a lys-Sepharose 4b column (10mL), and eluting an approximately kringle 1-3 fragment of plasminogen and other lysine binding fragments, such as kringle 4, with 5mL 200mM ϵ ACA/50mM Tris pH 7.4. The kringle 1-3 region protein was separated from kringle 4 and other smaller (<12kDa) fragments using gel filtration on a Superdex200 HR 10/30 column (Pharmacia). The Superdex200 column was equilibrated with PBS. Kringle 1-3 region protein concentration was determined spectrophotometrically at a wavelength of 280nm, using $A^{0.1\%/1cm}$ value of 1.87 (computed from the protparam tool available at <http://www.expasy.ch/> the ExPASy Molecular Biology World Wide Web (WWW) server at the Swiss Institute of Bioinformatics).

Bovine capillary endothelial cells were obtained as described previously in O'Reilly, M.S. *et al.* "Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma" *Cell* 79:315-328 (1994) and were maintained in DMEM with 10% heat-inactivated BCS, antibiotics, and 3ng/mL recombinant human bFGF (Scios Nova, CA). Cells were washed with PBS and dispersed in a 0.05 % solution of trypsin. A cell suspension was made with DMEM/10% BCS/1% antibiotics and the concentration adjusted to 25,000 cells/ml. Cells were plated onto gelatinized 24-well culture plates (0.5mL/well) and were incubated at 37°C in 10% CO₂ for 24 hours. The media was replaced with 0.25mL of DMEM/5% BCS/1% antibiotics, and the test sample applied.

After 20 minutes incubation, media and bFGF were added to each well to obtain a final volume of 0.5mL DMEM/5% BCS/1% antibiotics/1 ng/mL bFGF. After 72 hours, cells were dispersed in trypsin, resuspended in Hematell (Fisher scientific, PA) and counted by Coulter counter. Data from endothelial cell proliferation assays were plotted and IC₅₀ values determined using SigmaPlot. All data points were determined in triplicate from four separate preparations of kringle 4-51-3 region protein.

Example 2

Deglycosylation of plasminogen results in an increased yield of kringle 4-51-3 region proteins

Equal amounts of plasminogen 1 and plasminogen 2 were digested with PPE, and the kringle 4-51-3 region proteins purified as described in Example 1 using a combination of affinity chromatography (lys-Sepharose) to purify kringle 4-51-3 region proteins and other lysine binding fragments of plasminogen, and gel filtration (Superdex200) to separate kringle 4-51-3 region proteins from kringle 4. Figure 2 shows a typical chromatogram from the affinity column, demonstrating the reduced yield obtained when plasminogen 1 is used, as a substrate compared to plasminogen 2. Figure 3 shows the chromatogram obtained from a typical gel filtration experiment as described in Example 1). It was noted that kringle 4-51-3 region protein from plasminogen 1 has a slightly lower retention time (13.77 minutes) than deglycosylated kringle 4-51-3 region protein (14.36 minutes), as would be expected from this glycosylated fragment. The yield of kringle 4-51-3 region proteins from plasminogen 2 was 4-5 fold higher than the yield of kringle 4-51-3 region proteins using plasminogen 1 as a substrate. Glycosylated kringle 4-51-3 region proteins migrated at approximately 40 kDa and deglycosylated kringle 4-51-3 region proteins migrated at 38kDa on SDS-PAGE (data not shown). Edman degradation amino-terminal sequence analysis

of both kringle 1-51-3 region protein glycoforms, gave identical sequence starting at Arg87, 16 residues amino-terminal to the cysteine residue that marks the beginning of kringle 1.

5 Therefore, it was determined that the presence of
N-linked carbohydrate on Asn-289 modulates the generation of
kringle 1-51-3 region proteins from human plasminogen. The
presence of carbohydrate results in a 4-5-fold lower yield of
kringle 1-51-3 region protein when PPE is used to cleave
kringle 1-51-3 region proteins out of plasminogen. It was also
10 determined that fractionation of kringle 1-51-3 region proteins
generated from material containing glycosylated plasminogen
(plasminogen 1) and deglycosylated plasminogen (plasminogen
2) in the ratio 40:60 using lectin affinity chromatography
resulted in only 5-10% of glycosylated kringle 1-51-3 region
15 proteins (data not shown). This data further indicated that
N-linked carbohydrate is a negative modulator of kringle 1-51-3
region protein generation. Both kringle 1-51-3 region glycoforms
had identical N-terminal sequence, indicating that the
carbohydrate did not interfere with the site of proteolytic
20 cleavage.

Although glycosylated kringle 1-51-3 region proteins
were less efficiently generated and less efficient as an inhibitor
or angiogenesis, it may be that kringle 1-51-3 region proteins
containing N-linked carbohydrate, will have a longer clearance
25 time than deglycosylated kringle 1-51-3 region proteins. This
may be a mechanism to ensure persistence of anti-endothelial
activity systemically, albeit at a reduced level.

Example 3

30 *Deglycosylated kringle 1-51-3 region proteins have increased
antiangiogenic activity as compared to glycosylated kringle 1-
51-3 region proteins*

Both kringle 1-51-3 region protein glycoforms were
assayed for their ability to inhibit the proliferation of bovine
35 endothelial cells as described in Example 1. Both kringle 1-51-

3 region protein glycoforms inhibited endothelial cell proliferation. However, glycosylated kringle 4-51-3 region protein, which contains an N-linked carbohydrate, has a measured IC₅₀ value of approximately 13μg/mL, whereas
5 deglycosylated kringle 4-51-3 region protein, the glycoform, lacking an N-linked carbohydrate, has a measured IC₅₀ value of approximately 2.4μg/mL. Thus, deglycosylated kringle 4-51-3 region protein appears to be a much more efficient inhibitor of endothelial cell proliferation, and in particular was 4-5 fold
10 more efficient as an inhibitor of endothelial cell proliferation.

All patents, publications and abstracts cited above are hereby incorporated by reference. It should be understood that the foregoing relates only to preferred embodiments of the
15 present invention, and that numerous modifications or alterations may be made therein without departing from the spirit and the scope of the invention as set forth in the appended claims.

**DEGLYCOSYLATED KRINGLE 4-51-3 REGION
FRAGMENTS OF PLASMINOGEN AND
METHODS OF USE**

5

Abstract of the Invention

Disclosed are deglycosylated fragments of a kringle 4-51-3
region of plasminogen, nucleotides encoding deglycosylated
kringle 4-51-3 region proteins and antibodies specific for
10 deglycosylated kringle 4-51-3 region proteins. The
compositions of the present invention have increased
antiangiogenic activity as compared to previously isolated
kringle 4-51-3 region proteins. Also included in the present
invention are methods of treating angiogenesis-associated
15 diseases and conditions such as cancer using the compositions
described herein.

Attorney Docket: 05940-0141 (05213-2141, 43170-219913)

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